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Michael G. Kippenhan

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A NEW SPECIES OF *CORTICOTOMUS* SHARP, 1891 (COLEOPTERA: TROGOSSITIDAE: TROGOSSITINAE) FROM BRAZIL

MICHAEL G. KIPPENHAN
Montana Entomology Collection
Montana State University, Bozeman, MT 59717, USA
kippenhan138@gmail.com

ABSTRACT

Corticotomus feisorum Kippenhan, new species is described and illustrated from specimens collected in the Brazilian states of Rondônia and São Paulo; this new species is differentiated from other *Corticotomus* Sharp, 1891 species exhibiting similar dorsal coloration. Photos, a diagnosis, and new country records are provided for *Corticotomus quadrimaculatus* (Léveillé, 1894), a similarly colored species also occurring in Brazil. A key to the species of the genus inhabiting South America is provided.

Keywords: Cleroidea, smooth bark-gnawing beetles, South America, taxonomy

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INTRODUCTION

As currently recognized, the New World genus *Corticotomus* Sharp, 1891 is comprised of 15 species distributed from southern Canada south to Brazil and Chile, and the West Indies (Barron 1971; Kolibáč 2013; Lepesme 1947; Sharp 1891). Unlike other genera with similar distribution patterns in the New World—*Airora* Reitter, 1876, *Nemozoma* Latreille, 1804, *Temnoscheila* Westwood, 1830, and *Tenebroides* Piller and Mitterpacher, 1783—*Corticotomus* is unique in that it is most speciose in the Northern Hemisphere. Pending a comprehensive review of the genus to delineate species distribution, a survey of type localities indicates that only three of the 15 species have been described from South America (Kolibáč 2013)—*Corticotomus bicolor* (Léveillé, 1895) (type locality: Cochabamba, Bolivia), *Corticotomus fulvus* (Léveillé, 1905) (type locality: Minas Gerais, Brazil), and *Corticotomus quadrimaculatus* (Léveillé, 1894) (type locality: Bahia, Brazil).

Although adult specimens of the 16 species (after this study) comprising the current concept of *Corticotomus* are generally distinguishable, the boundaries of the genus warrant further investigation. The inclusion of species originally described in four other genera is reflected in the diversity of morphological characters employed in historical generic diagnoses. For example, although Sharp's (1891) original description accurately reflected the two species constituting his generic concept, he did not note that both possess a subhumeral impression on the elytron. By contrast, Barron's (1971) broader

generic concept encompassed the morphological diversity of North American species north of Mexico and emphasized the presence of the elytral subhumeral impression as a diagnostic generic character. Although convenient due to its apparent uniqueness among New World TrogoSSitidae, the elytron with a subhumeral impression is not present in all *Corticotomus* species and therefore cannot be used for generic-level diagnosis in its current constitution. Moreover, the works of both Sharp (1891) and Barron (1971) were based on geographically limited faunal studies which artificially narrowed generic delimitations. These generic boundaries were further obscured by Kolibáč's (2005) synonymization of *Colydobius* Sharp, 1891 and *Leveillesoma* Lepesme and Paulian, 1944 with *Corticotomus*, without explanation.

Generic identification can be further complicated by the observation that many of the diagnostic morphological traits of *Corticotomus* are shared, at least in part, with *Nemozoma*, *Euschaefferia* Leng, 1920, and similarly sized species of *Airora*. Accordingly, and for the purposes of the present work, adult members of *Corticotomus* are defined using readily observable morphological characters as follows: body very small to small (< 7 mm), relatively parallel-sided, elongate, cylindrical to subdepressed; pronotum and elytra subequal in width; head with frons flat to slightly concave, never bearing a longitudinal sulcus or conical depression; mandibles porrect to upturned, with the terminal and second teeth widely separated and the second tooth small; in lateral view, anterior margin of prothorax narrowly discontinuous, slightly, obtusely offset between dorsal and

ventral surfaces, never abruptly, acutely interrupted by projecting apical angles, pronotum with a thin lateral bead and lacking anterior or basal angles; antennal club loose, with antennomere 11 elongate-trapezoidal (Fig. 2B); protibia bearing a long, curved apical spur and up to two very short lateral spines (Fig. 2A). Although the generic redescrptions in Kolibáč (2005, 2013) are based on the type species and morphologically similar species, and thus include only the most generalized features, his key to the genera of Trogossitini (Kolibáč 2013) remains sufficient to distinguish *Corticotomus* from other trogossitid genera of similar body form (*i.e.*, very small to small, subdepressed to subcylindrical) occurring in the Neotropical region.

All *Corticotomus* species exhibit nonmetallic body coloration, ranging from uniformly ochraceous to piceous. Several species, characterized by a castaneous to piceous coloration, display bicolored elytra with the secondary contrasting color patch varying from ochraceous to rufous. Among these bicolored species, all but three—*Corticotomus gracilis* Sharp, 1891 (type locality: Bugaba, Panama), *Corticotomus signatus* (Sharp, 1891) (type locality: Volcan de Atitlan, Guatemala), and *C. quadrimaculatus*—have the contrasting color confined to a single, large area, either covering the basal or apical portion of the elytron. In contrast, the coloration of *C. gracilis*, *C. signatus*, and *C. quadrimaculatus* consists of two distinct color patches on each elytron, one in the basal region and the other in the apical region.

Little additional information has been published on the Neotropical species since their original descriptions apart from Kolibáč (2014), who included images and confirmed the generic affiliation of specimens he identified as *C. quadrimaculatus*, which were reared from the branches of *Anadenanthera falcata* (Benth.) Speg. (Fabaceae) from the state of São Paulo, Brazil. While identifying trogossitid specimens for the Florida State Collection of Arthropods, I identified two specimens matching the photos of *C. quadrimaculatus sensu* Kolibáč (2014), including one labeled as belonging to the same series. Originally placed in the genus *Airora*, Léveillé later transferred this species to *Corticotomus*, commenting on its resemblance to *C. gracilis* (Léveillé 1905). In 2024, during a visit to the Muséum National d'Histoire Naturelle, Paris, I examined of the type of *Airora quadrimaculata* (Fig. 4), at which time it became apparent that the specimen identified as *C. quadrimaculatus* by Kolibáč (2014) was not, in fact, conspecific with that species and instead represented an undescribed species. This new species is described below.

MATERIALS AND METHODS

Specimens from the following collections were studied:

CASC	California Academy of Sciences, San Francisco, CA, USA
CNCI	Canadian National Collection of Insects, Arachnids and Nematodes, Ottawa, Canada
FSCA	Florida State Collection of Arthropods, Gainesville, FL, USA
MGKC	Michael G. Kippenhan Collection, McMinnville, OR, USA
MNHN	Muséum National d'Histoire Naturelle, Paris, France
NHMUK	Natural History Museum, London, UK

This study is based on the examination of the primary types for the following *Corticotomus* taxa (following Kolibáč 2013): *Corticotomus apicalis* Van Dyke, 1944 (CASC), *Corticotomus basalis* Sharp, 1891 (NHMUK), *C. bicolor* (MNHN), *Corticotomus californicus* Van Dyke, 1915 (CASC), *Corticotomus divisus* (Sharp, 1891) (NHMUK), *Corticotomus dufau* (Léveillé, 1907) (MNHN), *C. fulvus* (MNHN), *C. gracilis* (NHMUK; note: this species was omitted in Kolibáč 2013), *C. quadrimaculatus* (MNHN), *Corticotomus sharpi* Léveillé, 1905 (MNHN), *C. signatus* (NHMUK). The taxonomic concepts of *Corticotomus caviceps* (Fall, 1910) (<https://mczbase.mcz.harvard.edu/guid/MCZ:Ent:24477>) and *Corticotomus cylindricus* (LeConte, 1863) (<https://mczbase.mcz.harvard.edu/guid/MCZ:Ent:7035>) are based on type specimens housed at the Museum of Comparative Zoology, Cambridge, MA, USA, redescrptions by Barron (1971), and specimens housed in MGKC. The taxonomic concepts of *Corticotomus parallelus* (Melsheimer, 1844) and *Corticotomus depressus* Schaeffer, 1918 are based on redescrptions by Barron (1971) and specimens housed in MGKC.

Measurements were taken with a Bioquip 5-mm hand-held micro-ruler; measurements for the head, pronotum, and elytra were taken independently: head length (HL) was measured from the tip of the frontoclypeus to the anterior margin of the pronotum; head width (HW) was the maximum width across the eyes; pronotal length (PL) was measured along the midline from the anterior to posterior margins; pronotal width was measured across the maximum width; elytral length (EL) was measured from the humeral angle to the apex; elytral width (EW) was measured across both elytra at the maximum width. Quoted specimen label data utilizes a forward slash (/) to represent line breaks and a double forward slash (//) to separate labels. All data are

presented verbatim. All labels are typeset unless otherwise noted. Dissection and illustration techniques follow Kippenhan (2025).

TAXONOMY

Corticotomus feisorum Kippenhan, new species

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(Figs. 1–3)

Diagnosis. This species is identifiable based on the following character combination: robust, subcylindrical body, dorsal surfaces piceous to castaneous with two large rufous to testaceous patches on each elytron, slightly upturned mandibles, elytron lacking defined striae with punctures appearing unordered and all punctures subequal in diameter, elytron lacking subhumeral impression, with basal marginal bead extending obliquely towards suture, and submentum with denticles large and interdenticular spaces narrow and

strongly concave. The other three *Corticotomus* species with two contrasting color patches on each elytron—*C. gracilis*, *C. signatus*, and *C. quadrimaculatus*—all have elytral punctation arranged in definitive striae with the punctures in the interstriae intervals being approximately half the diameter of the striae punctures, subhumeral elytral impressions, and the submentum with small denticles and central interdenticular space broad and shallowly concave. Of these three species, only *C. gracilis* shares the robust, subcylindrical body and slightly upturned mandibles as *C. feisorum*.

Type Specimens. **Holotype:** Male, “24/III/2011 / BR-SP-Ilha Solteira / UNESP campus / 20°25’11.65” S 51°20’28.19” W / ex broken *Anadenanthera fal-* / *cata* limb attacked by *Pseu-* / *dochramesus acteclavatus* Flechtmann, C.A.H. col. // [red label] HOLOTYPE / *Corticotomus feisorum* / M. Kippenhan” (FSCA). **Paratype** (*n* = 1): Sex undetermined, “BRAZIL: Rondônia 62 / km SW Ariquemes, nr / Fzsa. Rancho Grande / 8-20-XI-1994 JE Eger /



Fig. 1. *Corticotomus feisorum*, new species, holotype (FSCA). A) Dorsal habitus (bm = basal marginal bead), B) Lateral habitus. Scale bar is for A and B.

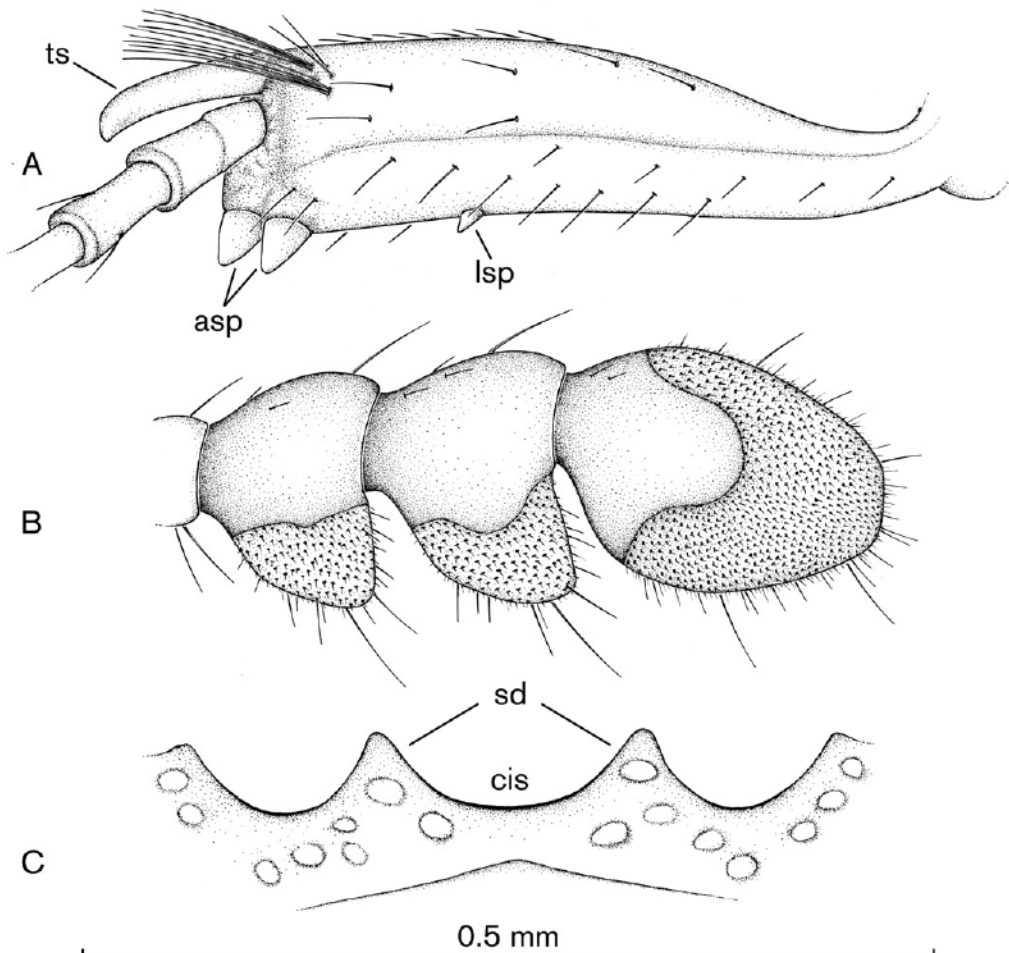


Fig. 2. Adult structures of *Corticotomus feisorum*, new species. A) Left protibia, anterolateral view (ts = tibial spur, asp = apical spine, lsp = lateral spine), B) Antennal club, C) Submentum (sd = submentum denticle, cis = submentum interdenticle space). Scale bar is for A–C.

Black Light Trap // [yellow label] PARATYPE / *Corticotomus feisorum* / M. Kippenhan" (FSCA).

Description. Body small, robust, subcylindrical, overall length: 2.8–3.4 mm; HL: 0.4–0.6 mm, HW: 0.65–0.8 mm, PL: 0.85–1.0 mm, PW: 0.72–0.9 mm, EL: 1.5–1.9 mm, EW: 0.8–1.0 mm. **Head** short, piceous with frons castaneous, relatively shiny; surface relatively smooth, with faint microsculpture; frontoclypeus with denticles widely spaced, well developed, subacute, interdenticle space strongly concave, anterolateral corners slightly produced, with denticles slightly extended; frons shallowly depressed, vertex relatively flat; dorsal surface with small, well impressed, longitudinal, subovular punctures relatively uniformly spaced approximately 1.0–1.5× diameter apart, slightly directed inward, punctation largest and deepest on vertex, gradually

becoming smaller and shallower towards frontoclypeal margin, frontoclypeal margin impunctate, temporal punctation small, circular, well impressed; gena with surface slightly irregular, with a few scattered, medium-sized, shallowly impressed punctures; submentum anterior margin with denticulation large, well developed (Fig. 2C, sd), central and lateral interdenticle spaces narrow, strongly concave (Fig. 2C, cis), posterior margin directed anteriorly, very slightly elevated; gena with a single, very long sensory seta; interocular distance broad, eyes moderate in size, slightly extended dorsally, slightly protuberant. **Mouthparts** with mentum unobserved; mandibles short, stout, slightly upturned; terminal labial and maxillary palpomeres longer than wide, fusiform, with narrow, truncate apex. **Antenna** long, extending slightly beyond anterior margin of

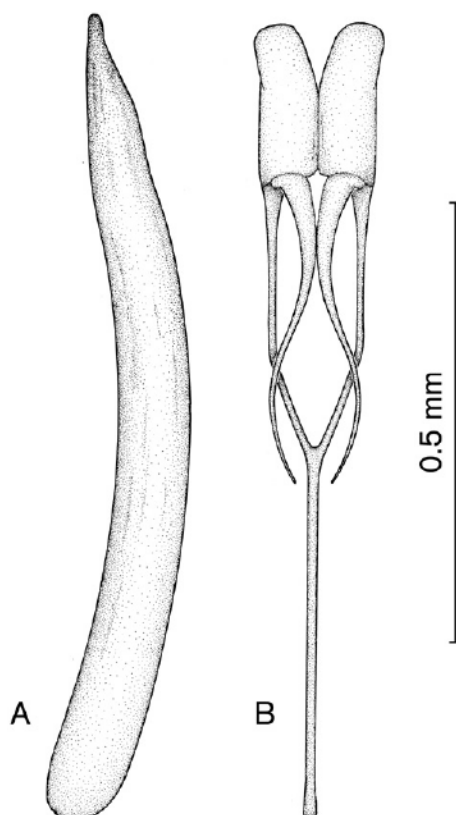


Fig. 3. Male genitalia of *Corticotomus feisorum*, new species. A) Median lobe, B) Tegmen. Scale bar is for A and B.

pronotum, scape large, globular, pedicel small, short, antennomeres 3–8 gradually longer and wider from 3 to 8, antennomeres 9–11 forming large, unilaterally expanded club, antennomeres 9 and 10 each with sensorial field covering approximately 40% of surface, antennomere 11 elongate-subtrapezoidal, sensorial field covering approximately 55% of surface (Fig. 2B), antennomeres with base castaneous, becoming rufous to testaceous at apex. **Pronotum** piceous to castaneous, relatively shiny; surface smooth, with very faint microsculpture; robust, somewhat bulbous, subrectangular, sides shallowly convex, in dorsal view anterior margin wide, relatively straight with apical angles narrowly, shallowly convex, not produced, basal margin short, relatively straight with apices broadly, obtusely rounded, in lateral view anterior margin of prothorax narrowly discontinuous, slightly, obtusely offset between dorsal and ventral surfaces, with complete, thin lateral bead; disc with subcircular, small, deeply impressed, relatively uniformly spaced punctures, separated by 1.0–2.0 $\times$ diameter, punctures along margins smaller

and more shallowly impressed, punctures bordering lateral bead deeply, widely impressed; hypomeron impunctate. **Scutellar shield** short, triangular. **Elytra** strongly convex with apical margin declivous, basal third castaneous to rufous, middle third piceous, apical third piceous with longitudinally directed, subovular, castaneous to testaceous patch located closer to suture than lateral margin, suture castaneous, shiny; surface relatively smooth, with very faint microsculpture; sides slightly convex, widest in apical three-quarters, apex obtusely rounded; punctures circular, smaller than pronotum, well impressed, irregularly spaced, approximately 1.5–2.5 $\times$ diameter apart, punctures on lateral surfaces less deeply impressed; basal marginal bead well defined on humerus, raised, extending obliquely towards suture (Fig. 1A, bm). **Prosternum** castaneous to rufous, shiny; surface glabrous, with very faint microsculpture; with medium-sized to small, shallowly to faintly impressed, uniformly widely spaced, round punctures throughout, anterior margin broadly impunctate; in lateral view intercoxal process with apex declivous, broadly convex, surface with a few small, moderately impressed, widely spaced punctures. **Mesoventrite** castaneous to rufous, shiny; surface glabrous, with faint microsculpture; medially impunctate, lateral area with small, well-impressed punctures. **Metaventrte** castaneous to rufous, shiny; surface glabrous, with faint microsculpture; with a few widely spaced, small, well-impressed punctures throughout. **Legs** castaneous to rufous, shiny; profemur broad, protibia broadly expanded at apex, outer margin relatively straight, apical spur longer than tarsomeres 1 + 2 (Fig. 2A, ts), with 2 rearward-facing apical spines (Fig. 2A, asp), 1 very small lateral spine (Fig. 2A, lsp); meso- and metatibiae each with short, sublinear apical spur, without lateral spines, apex less broadly expanded. **Abdomen** castaneous to rufous, shiny; ventrites glabrous, with very faint microsculpture; ventrites small, with shallowly impressed punctures in basal half. **Aedeagus** with median lobe subequal in length to tegmen, narrow, moderately convex, apex elongated to ovoid tip (Fig. 3A); tegmen with apical lobes short, approximately 0.18 $\times$ length of tegmen, basal portion fused, elongate, approximately 0.45 $\times$ length of tegmen, setae unobserved (Fig. 3B).

Variation. The paratype is lighter in color than the holotype.

Etymology. This species is named in honor of the type locality—Faculdade de Engenharia de Ilha Solteira (FEIS)—a campus of the São Paulo State University Júlio de Mesquita Filho (UNESP).

Distribution. The two localities represented by the type series are approximately 1,700 km apart in the states of Rondônia and São Paulo, Brazil.

Remarks. Kolibáč (2014) reported that specimens of this species (referred to there as *C. quadrimaculatus*) were reared from felled branches of *A. falcata* that were later attacked by *Pseudochramesus acuticlavatus* Hagedorn (Curculionidae: Scolytinae), as reported by Flechtmann and Costa (2012).

***Corticotomus quadrimaculatus* (Léveillé, 1894)**
(Fig. 4)

Airora quadrimaculata Léveillé 1894: CXLVI.
Holotype: “[green label, manuscript] Bahia / Brazil // [handscript] 4 • maculatus Lév // 4-maculatus / Lév. / Brasilia // MUSEUM PARIS / COLL. A. LÉVEILLÉ // [red label] Type. // [red label] HOLOTYPE // HOLOTYPE / *Airora* /

quadrimaculata Léveillé, / 1894 // MNHN, Paris / EC25288 [QR code]”. Type locality: “Brésil: Bahia”. Type repository: MNHN (examined).
Corticotomus quadrimaculatus: Léveillé 1905: 30; Léveillé 1910: 7; Blackwelder 1944: 392; Kolibáč 2013: 56.

Diagnosis. Body narrow, subparallel, subdepressed, dorsal surfaces piceous with two small, rufous patches on each elytron, head with mandibles porrect, head and pronotum with small, well-impressed, densely, relatively uniformly spaced punctures, elytra with punctures arranged into striae, punctures becoming less impressed apically, basal margin defined on humerus only, with shallow, short subhumeral impression.

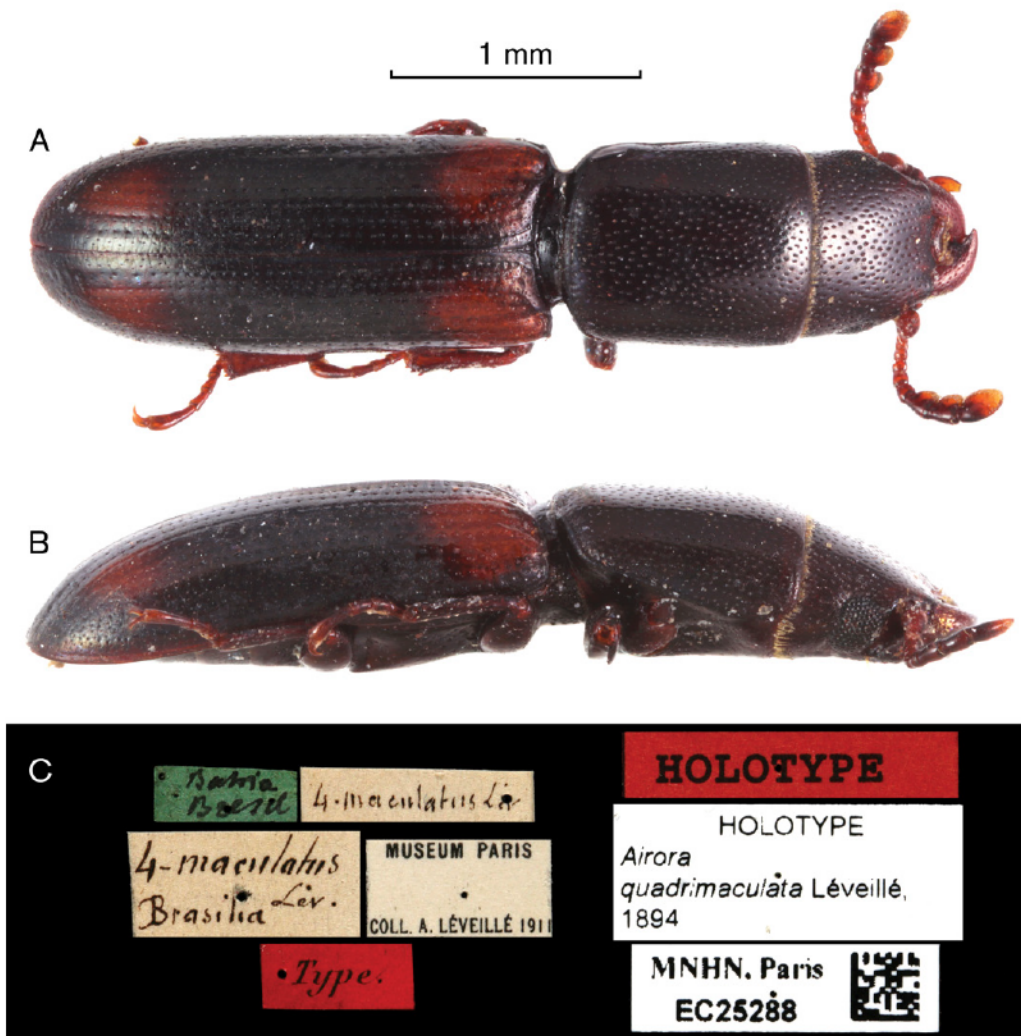


Fig. 4. *Airora quadrimaculata*, holotype (MNHN). A) Dorsal habitus, B) Lateral habitus, C) Labels. Scale bar is for A and B.

Specimens Examined ($n = 8$). **Bolivia:** Santa Cruz, El Refugio Los Volcanes, 18°06.364'S 63°35.554'W, 4–9-X-2007, elev. 3,363 ft., R. Morris, at light (1 FSCA; 1 MGKC). **Brazil:** Nova Teutonia, 27°11'S 52°23'W, 300–500 m, XII-1972, Fritz Plaumann (1 CNCI). **Peru:** San Martín, Pumarinri Lodge, 10 km E. Shapaja, 24–27-X-2012, J. B. Heppner, 225 m (1 FSCA); San Martín, Moyabamba, vic. Ecologico Rumipata, 13–18-X-2012, J. E. Eger (1 FSCA). **Trinidad:** Simla, Arima-Blanchisseuse Rd., 11-VII-1975, J. Price, blacklight trap (2 FSCA). **Venezuela:** Falcón, nr. Piritu, 27-VII-1993, B. K. Dozier (1 FSCA). Bolivia, Peru, Trinidad and Tobago, and Venezuela represent **new country records**.

Remarks. Previously only recorded from Brazil (Blackwelder 1944; Léveillé 1910), the new records above indicate this species is widely distributed in South America.

Key to the South American Species of *Corticotomus*

1. Antennomeres 3 and 4 elongate, narrow, antennomeres 9 and 10 asymmetrically bilaterally dilated, with sensorial fields on each side; body unicolored, ochraceous *C. fubvus* (Léveillé)
- 1'. Antennomeres 3 and 4 subequal in form to antennomeres 5–8, antennomeres 9 and 10 unilaterally dilated; body castaneous to piceous with 1 or 2 contrasting lighter patches on each elytron 2
2. Elytral punctation unordered (Fig. 1A); elytron without subhumeral impression (Figs. 1A–B); mandibles slightly upturned (Fig. 1B) *C. feisorum* Kippenhan, new species
- 2'. Elytral punctation serially arranged (Fig. 4A); elytron with subhumeral impression (Figs. 4A–B); mandibles porrect (Fig. 4B) 3
3. Elytron with 2 contrasting lighter color patches, 1 basal, 1 apical (Figs. 4A–B) *C. quadrimaculatus* (Léveillé)
- 3'. Elytron with contrasting, lighter color patches restricted to basal third *C. bicolor* (Léveillé)

DISCUSSION

The large geographic gap between the two specimens of *C. feisorum*, along with the fact that it is one of only four *Corticotomus* species recorded from South America, raises questions about whether the distribution pattern of this genus reflects its true biogeography or is simply an artifact of limited collecting efforts. Considering that *C. quadrimaculatus* was previously only recorded from Brazil (Blackwelder

1944; Léveillé 1910), the four new country records indicate that this species is more widely distributed across South America, thus supporting the idea that current biogeographical knowledge of the genus may be biased by historical gaps in sampling and regional underrepresentation.

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