

Imidazo[1,2-a]Pyridine-3-Carboxamides Are Active Antimicrobial Agents against Mycobacterium avium Infection In Vivo

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Abstract
A panel of six imidazo[1,2-a]pyridine-3-carboxamides (IAPs) were shown to have low micromolar activity
against <i>Mycobacterium avium</i> strains. Compound ND-10885 (2) , showed significant activity in the lung,
spleen and liver in a mouse <i>M. avium</i> infection model. A combined regimen consisting of ND-10885 (2)
and rifampin were additive in their anti-M. avium activity in the lung. Our data indicates that IAPs
represent a new class of antibiotics that are active against <i>M. avium</i> and could potentially serve as an
effective addition to a combined treatment regimen.

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States (1) (2). Mycobacterium avium complex (MAC), which consists of M. avium and M. intracellulare, is an important cause of both pulmonary disease in individuals with underlying lung diseases such as cystic fibrosis and chronic obstructive pulmonary disease and is an opportunistic pathogen in immunocompromised patients (3, 4). Among the NTM species isolated from U.S. patients, 80% were classified as MAC (5). MAC is ubiquitous within the environment and is found in soil, treated or untreated water, house plumbing systems and animals (6). MAC infection is difficult to treat and has been shown to be resistant to many of the clinically used anti-tuberculosis agents (7, 8). We previously disclosed a novel family of compounds, imidazo[1,2-a]pyridine-3-carboxamides (IAPs), with potent activity against M. tuberculosis (Mtb) (9-12). The mechanism of action and anti-Mtb in vivo efficacy of this exciting new class has been documented by us and other groups (9, 13-16). Through a "hit" to "lead" optimization effort aided by the Lilly TB Drug Discovery Initiative (LTBDDI) additional IAP compounds (1-6) were generated and found to have encouraging in vivo pharmacokinetics (PK). Herein, we describe activity of these latest analogs against M. avium both in vitro and in vivo. Activity of selected compounds in vitro. Six compounds having diverse PKs were selected and synthesized following our published methods (9-11). Experimental data and information on all previously uncharacterized compounds (1, 2 and 6) can be found in the supporting information section. MIC studies were performed using a resazurin-based colorimetric assay and CFU quantification as described (17). Screening the IAPS against M. avium strains 101 and 2151 using standard protocols indicated that they had moderate potency (Table 1). The activity of these compounds against M. avium $(2.6 - 27.8 \,\mu\text{M}, \text{two strains})$ was limited relative to Mtb (Table S1) but comparable to positive controls of clarithromycin and azithromycin (1.4 and 13.4 µM, respectively). These compounds also had a good

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The incidence of non-tuberculosis mycobacteria (NTM) infections has been increasing in the United

therapeutic window when screened against VERO cells (9) (Table S1).

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All six compounds were evaluated for their ability to kill or inhibit *M. avium* replication *in vitro*. Five out of six compounds were bactericidal or bacteriostatic (Fig 2). Consistent with its MIC value, ND-9903 (5) showed no activity against M. avium 101 at the highest concentration tested. Separate studies of ND-9758 (3), ND-9759 (4) and ND-10890 (6) at a 1.0 μ g/ml each indicated that they were bacteriostatic as the bacterial counts were maintained near the original inoculum. ND-10885 (2) and ND-9873 (1) had bactericidal activity against M. avium 101 at 1.0 µg/ml as bacterial numbers decreased by 1 log₁₀ and 0.5 log₁₀, respectively, compared to the original inoculum. Rifampin was the positive control and showed the best bactericidal activity against M. avium 101. Except for ND9903 (9), all compounds showed dosedependent activity against M. avium. ND-10885 (2) also showed activity against various other M. avium clinical isolates of different serotypes, although again it varied 10 fold or more between strains. Pharmacokinetics. Single-dose pharmacokinetics of compounds 1 - 6 were determined in uninfected 8week-old male Balb/c mice. Mice received by oral gavage a single dose of compound 1 (at 10 and 100 mg/kg), compound 2 (at 100 mg/kg), compound 3 (at 10 mg/kg), compound 4 (at 30 mg/kg), compound 5 (at 100 mg/kg), and compound 6 (at 10 and 100 mg/kg). Compounds were analyzed as previously described (10). Calculated parameters include clearance (CI), area-under-the-curve (AUC), half-life (T1/2), maximum serum concentration (Cmax), time of maximum concentration (Tmax), bioavailability (%F) and percent drug fraction unbound in plasma (Fu, Plasma). As shown in table 2, all of these compounds had high plasma exposure at their respective doses but there was a large range in half-lives observed (from 2.3 to >24 h). In our drug development paradigm, we selected the maximum free drug concentration per dose (Cmax, u) as the most meaningful pharmacokinetic property to use for compound advancement and ND-10885 (2) and ND-10890 (6) had the highest values (2,795 and 4,232 nM, respectively). The free drug concentration for those two compounds were high enough above their MICs that they were anticipated to be effective against MAC 101 in vivo. The PK parameters for compounds 1, 2 and 6 by IV dose can be found in Table S3.

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(Log₁₀) in the lung.

Maximum tolerated dose. Five compounds were evaluated in mice to determine their maximum tolerated dose as previously described (13). Compounds ND-9873 (1) and ND-10885 (2) were tolerated at the highest concentration of 250 mg/kg for one week, ND-10890 (6) was well-tolerated at 100 mg/kg for one week and as previously reported ND-9759 (5) was tolerated at 30 mg/kg for 28 days (13). Mice treated with ND-9758 (3) did not tolerate the drug even at the lowest concentration (30 mg/kg). Efficacy of ND10885 in MAC-infected mice. We chose compound ND-10885 (2) for the in vivo studies in Wild type Balb/c mice based on its relatively good in vitro bactericidal activity against M. avium, its PK profile and its low toxicity in mice. Wild type Balb/c mice (n=3) were retro-orbitally infected with M. avium MAC 101 at a dose of 10^7 CFU in 50 μ L of PBS (18). One week after the infection, mice were treated by oral gavage with ND-10885 (2) dissolved in 80% propylene glycol (v/v) once daily 6 days a week for two weeks. After the final dosing, all mice were sacrificed, and the mycobacaterial burden was determined as described previously (13). The bacterial numbers were quantified by visually counting bacterial colonies. At the beginning of treatment, a group of mice (n=3) were sacrificed to measure the mycobacterial input in the lung, spleen and liver. As a negative control, a group of mice (n=3) were treated with the vehicle, 80% propylene glycol, only. ND-10885 (2) significantly inhibited M. avium growth in the lungs, spleens and livers when compared to the vehicle-treated M. avium-infected mice (Fig 3). The inhibitory activity of ND-10885 (2) was comparable to that of rifampin in all three organs. In addition, compared to single compound/drug regimens, mycobacterial counts were lower in all three organs when M. avium-infected mice were treated with a combined regimen of ND-10885 (2) and rifampin, although this was only statistically significant in the lung. When compared with baseline mycobacterial counts at the initiation of treatment, the combined regimen showed bactericidal activity with the CFU count decreasing 1.5-fold

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In conclusion, we show that IAPs are active against M. avium clinical isolates. One compound, ND-10885 (2), has significant activity against M. avium in mice, reducing bacterial burden in the lung, spleen and liver when compared to the untreated group. Most interestingly, ND-10885 (2) has activity comparable to rifampin, a critical component in the combined regimen used to treat MAC infections (8) and a combined regimen consisting of ND-10885 (2) and rifampin showed enhanced bactericidal activity in the lung. Our data suggest that IAPs should be pursued as a new class of compounds to treat M. avium and perhaps other NTMs.

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4. Cowman S, Loebinger M. 2015. Nontuberculous Mycobacterial Pulmonary Disease. Clinical

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Pulmonary Medicine, 22(1), 8-14. 5. Prevots DR, Shaw PA, Strickland D, Jackson LA, Raebel MA, Blosky MA, Montes de Oca R, Shea YR, Seitz AE, Holland SM, Olivier KN. 2010. Nontuberculous Mycobacterial Lung Disease Prevalence at Four Integrated Health Care Delivery Systems. American Journal of Respiratory and Critical Care Medicine 182:970-976. 6. Halstrom S, Price P, Thomson R. 2015. Review: Environmental mycobacteria as a cause of human infection. International Journal of Mycobacteriology 4:81-91. 7. Wagner D, Young LS. 2004. Nontuberculous mycobacterial infections: a clinical review. Infection **32**:257-270. 8. Griffith DE, Aksamit T, Brown-Elliott BA, Catanzaro A, Daley C, Gordin F, Holland SM, Horsburgh R, Huitt G, Iademarco MF, Iseman M, Olivier K, Ruoss S, von Reyn CF, Wallace RJ, Winthrop K, ATS Mycobacterial Diseases Subcommittee, American Thoracic Society, Infectious Disease Society of America. 2007. An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. Am J Respir Crit Care Med 175:367-416. 9. Moraski GC, Markley LD, Hipskind PA, Boshoff H, Cho S, Franzblau SG, Miller MJ. 2011. Advent of Imidazo[1,2-a]pyridine-3-carboxamides with Potent Multi- and Extended Drug Resistant Antituberculosis Activity. ACS Med Chem Lett 2:466-470. 10. Moraski GC, Markley LD, Cramer J, Hipskind PA, Boshoff H, Bailey M, Alling T, Ollinger J, Parish T, Miller MJ. 2013. Advancement of Imidazo[1,2-a]pyridines with Improved Pharmacokinetics

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11. Moraski GC, Miller PA, Bailey MA, Ollinger J, Parish T, Boshoff HI, Cho S, Anderson JR,

and Nanomolar Activity Against Mycobacterium tuberculosis. ACS Med Chem Lett 4:675-679.

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- Mulugeta S, Franzblau SG, Miller MJ. 2015. Putting Tuberculosis (TB) To Rest: Transformation of the Sleep Aid, Ambien, and "Anagrams" Generated Potent Antituberculosis Agents. ACS Infect Dis 1:85-90.
- 12. Ollinger J, Bailey MA, Moraski GC, Casey A, Florio S, Alling T, Miller MJ, Parish T. 2013. A Dual Read-Out Assay to Evaluate the Potency of Compounds Active against Mycobacterium tuberculosis. PLoS ONE 8:e60531.
 - 13. Cheng Y, Moraski GC, Cramer J, Miller MJ, Schorey JS. 2014. Bactericidal Activity of an Imidazo[1, 2-a]pyridine Using a Mouse M. tuberculosis Infection Model. PLoS ONE 9:e87483.
- 170 14. Abrahams KA, Cox JAG, Spivey VL, Loman NJ, Pallen MJ, Constantinidou C, Fernández R, 171 Alemparte C, Remuiñán MJ, Barros D, Ballell L, Besra GS. 2012. Identification of Novel 172 Imidazo[1,2-a]pyridine Inhibitors Targeting M. tuberculosis QcrB. PLoS ONE 7:e52951.
- 173 15. Mak PA, Rao SPS, Ping Tan M, Lin X, Chyba J, Tay J, Ng SH, Tan BH, Cherian J, Duraiswamy J, 174 Bifani P, Lim V, Lee BH, Ling Ma N, Beer D, Thayalan P, Kuhen K, Chatterjee A, Supek F, Glynne 175 R, Zheng J, Boshoff HI, Barry CE, Dick T, Pethe K, Camacho LR. 2012. A High-Throughput Screen 176 To Identify Inhibitors of ATP Homeostasis in Non-replicating Mycobacterium tuberculosis. ACS 177 Chemical Biology 7:1190-1197.
 - 16. Pethe K, Bifani P, Jang J, Kang S, Park S, Ahn S, Jiricek J, Jung J, Jeon HK, Cechetto J, Christophe T, Lee H, Kempf M, Jackson M, Lenaerts AJ, Pham H, Jones V, Seo MJ, Kim YM, Seo M, Seo JJ, Park D, Ko Y, Choi I, Kim R, Kim SY, Lim S, Yim S-A, Nam J, Kang H, Kwon H, Oh C-T, Cho Y, Jang Y, Kim J, Chua A, Tan BH, Nanjundappa MB, Rao SPS, Barnes WS, Wintjens R, Walker JR, Alonso S, Lee S, Kim J, Oh S, Oh T, Nehrbass U, Han S-J, No Z, Lee J, Brodin P, Cho S-N, Nam K, Kim J. 2013. Discovery of Q203, a potent clinical candidate for the treatment of tuberculosis. Nat

184 Med 19:1157-1160. 17. Palomino J-C, Martin A, Camacho M, Guerra H, Swings J, Portaels F. 2002. Resazurin microtiter 185 186 assay plate: simple and inexpensive method for detection of drug resistance in Mycobacterium 187 tuberculosis. Antimicrob Agents Chemother 46:2720–2722. 188 18. Yardeni T, Eckhaus M, Morris HD, Huizing M, Hoogstraten-Miller S. 2011. Retro-orbital 189 injections in mice. Lab Anim (NY) 40(5) 155-60. 190

191 Table 1. Screening of compounds $\mathbf{1} - \mathbf{6}$ against two strains of M. avium. MICs are shown in μM .

Compound	Mol. Wt.	Clog P	MAC 101	MAC 2151
			(serotype 1)	(serotype 2)
ND-9873 (1)	363.34	4.6	2.8	27.5
ND-10885 (2)	321.38	3.6	1.6	15.6
ND-9758 (3)	389.43	5.8	1.28	2.57
ND-9759 (4)	405.88	6.4	0.31	1.2
ND-9903 (5)	425.41	6.4	>23.5	>23.5
ND-10890 (6)	382.44	3.4	2.6	13.1
Rifampin	822.95	6.04	0.077	0.158
Ethambutol	204.31	0.12	48.9	>48.9
Clarithromycin	747.95	2.82	1.4	1.4
Azithromycin	748.88	2.28	13.4	>13.4

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196 Table 2. Pharmacokinetic parameters of compounds 1-6.

	in vivo PK		
Compound	PK Value	PK calculation	
OCF ₃ ND-9873 (1)	AUC(ng*Hours/mL) =27,768 (PO; 10 mg/kg) T1/2 (Hrs)=2.91 F%=51 Cmax (ng/mL)= 10,294 Tmax (Hrs)=1.0 Cl (mL/Min./Kg)=20 Fu, PLasma= 0.023	- Cmax, u (100 mg/kg, PO) = 652 nM	
ND-10885 (2)	AUC(ng*Hours/mL) =51,249.7 (PO; 100 mg/kg) T1/2 (Hrs)=2.35 F%=44.3 Cmax (ng/mL)= 19,530 Tmax (Hrs)=0.333 Cl (mL/Min./Kg)=4.25 Fu, PLasma= 0.046	- Cmax, u (100 mg/kg, PO) = 2,795 nM	
ND-9758 (3)	AUC(ng*Hours/mL) =11,000 (PO; 10 mg/kg) T1/2 (Hrs)=13.2 F%=ND Cmax (ng/mL)= 1,160 Tmax (Hrs)=0.50 CI (mL/Min./kg)=NDA Fu, PLasma= 0.003	ND	
ND-9759 (4)	AUC(ng*Hours/mL) =22,200 (PO; 30 mg/kg) AUC T1/2 (Hrs) = 20.2 F% = ND Cmax (ng/mL) = 2,900 Tmax (Hrs) = 1.00 CI (mL/Min./kg)=NDA Fu, PLasma=0.001	- Cmax, u (30 mg/kg, PO) = 7 nM	
ND-9903 (5)	AUC(ng*Hours/mL) =594,000 (PO; 100 mg/kg) T1/2 (Hrs) > 24 F% = ND Cmax (ng/mL)= 34,900 Tmax (Hrs)=12 CI (mL/Min./kg)=NDA Fu, PLasma=0.0016	Cmax, u (100 mg/kg, PO) = 133 nM	
ND-10890 (6)	AUC(ng*Hours/mL) =32,800 (PO; 10 mg/kg) T1/2 (Hrs)=3.96 F%=46.3 Cmax (ng/mL)= 11,500 Tmax (Hrs)=0.250 CI (mL/Min./Kg)=2.32 Fu, PLasma=0.039	- Cmax, u (10 mg/kg, PO) = 1,232 nM - Cmax, u (100 mg/kg,PO) = 4,232 nM	

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198 ND; not determined

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Figure Legends
Fig. 1. Anti-M. avium activities of Imidazo[1,2-a]pyridine-3-carboxyamides in vitro measured by CFU
counts. M. avium were treated with compounds at various concentrations as shown, and then bacterial
CFU were determined on Middlebrook 7H10 agar plates. Baseline, M. avium CFU at the beginning of
treatment. The results are representatives of three independent experiments. *, p < 0.05 compared to
the Baseline (one-way ANOVA with Tukey post-test).
Fig. 2. Efficacy of ND10885 (6) against <i>M. avium</i> infection in the Balb/c mouse model. <i>M. avium</i> -
infected mice were treated with compounds once daily, 6 day per week for two weeks. Bacterial burder
in the lung, spleen and liver was determined. Mock, mice treated with vehicle alone. Baseline, bacterial
burden at the beginning of treatment. The results are representatives of two independent experiments.
*, p < 0.05 compared to the mock, ** p < 0.05 compared to ND10885 (100 mg/kg) (one-way ANOVA
with Tukey post-test).

Fig. 1

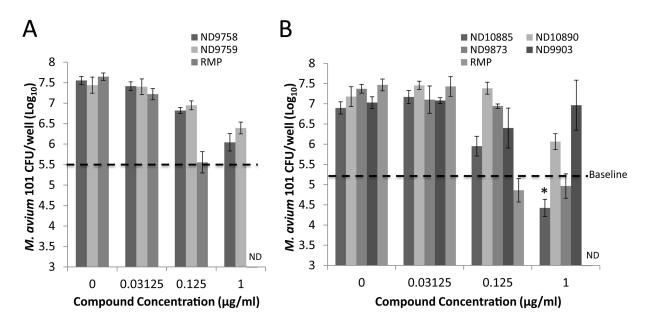


Fig. 2

