

# Genome Sequence of *Janthinobacterium* sp. CG23\_2, a Violacein-Producing Isolate from an Antarctic Supraglacial Stream

Heidi J. Smith,<sup>a</sup>  Christine M. Foreman,<sup>a</sup> Tatsuya Akiyama,<sup>a</sup> Michael J. Franklin,<sup>a</sup> Nicolas P. Devitt,<sup>b</sup> Thiruvarangan Ramaraj<sup>b</sup>

Center for Biofilm Engineering, Montana State University, Bozeman, Montana, USA<sup>a</sup>; National Center for Genome Resources (NCGR), Santa Fe, New Mexico, USA<sup>b</sup>

**Here, we present the draft genome sequence for the violacein-producing *Janthinobacterium* sp. CG23\_2 isolated from an Antarctic supraglacial stream. The genome is ~7.85 Mb, with a G+C content of 63.5%. The genome includes 7,247 candidate protein coding genes, which may provide insight into UV tolerance mechanisms.**

Received 22 October 2015 Accepted 7 December 2015 Published 28 January 2016

**Citation** Smith HJ, Foreman CM, Akiyama T, Franklin MJ, Devitt NP, Ramaraj T. 2016. Genome sequence of *Janthinobacterium* sp. CG23\_2, a violacein-producing isolate from an Antarctic supraglacial stream. *Genome Announc* 4(1):e01468-15. doi:10.1128/genomeA.01468-15.

**Copyright** © 2016 Smith et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Christine M. Foreman, cforeman@montana.edu.

Organisms inhabiting Antarctic supraglacial environments are subjected to a variety of environmental stresses. Ozone depletion over Antarctica has altered the spectral composition of solar radiation (1), and harmful UVC (100 to 280 nm) radiation is present on ice surfaces (2). Inescapable to microorganisms throughout the course of the austral summer, UV radiation damages biological macromolecules, including nucleic acids, lipids, and proteins and may lead to cell death. In order for microorganisms to inhabit environments with high UV radiation, efficient protective mechanisms and DNA and protein repair mechanisms are necessary. One mechanism for the protection from UV induced biological damage is for cells to produce protective pigments (3, 4).

The purple pigment violacein is produced by certain members of the  $\beta$ -proteobacteria, including some *Janthinobacterium* strains. Violacein has been investigated for its antimicrobial (5), antioxidant (6), and UV protection properties (4). Although the exact mechanism of violacein protection against UV damage is currently not well understood, evidence suggests that violacein can detoxify free radicals induced by UV radiation (4).

*Janthinobacterium* spp. are found in many different environments, including lakes, soils, and glaciers (7–9). We recently reported the genome sequence of a nonpigmented *Janthinobacterium* sp. isolated from a supraglacial stream. Here we present the genome sequence of a violacein producing strain *Janthinobacterium* sp. CG23\_2, isolated from the same system in Antarctica. Genomic analyses of these strains will offer insight into UV tolerance mechanisms from environmentally relevant isolates.

*Janthinobacterium* sp. strain CG23\_2 was isolated from the Cotton Glacier in the Antarctic Dry Valleys (77° 07S, 161° 50E). The organism was isolated on R2A agar medium incubated in the dark at 4°C for 12 days. *Janthinobacterium* sp. strain CG23\_2 is a psychrotolerant, aerobic, violacein-pigmented, rod-shaped, Gram-negative, catalase-positive organism. Genomic DNA was isolated following standard cetyltrimethylammonium bromide (CTAB) isolation protocols (<http://www.jgi.doe.gov>).

Whole-genome DNA sequencing was performed using a Pacific Biosciences (PacBio, Menlo Park, CA) RS II instrument (10). A single molecule real-time (SMRT) cell library was constructed with 10  $\mu$ g input DNA using the PacBio 20-kbps protocol. The library was then loaded onto one SMRT cell and sequenced using P5 polymerase and C3 chemistry with 180-min movie times. Sequencing yielded a total of 99,287 reads with mean read length of 5.9 kbps, totaling 581,513,481 bps ( $\approx$ 85-fold coverage). *De novo* assembly was constructed using the hierarchical genome assembly process (HGAP2) protocol from SMRT Analysis v2.0, including consensus polishing with Quiver (11, 12). The final assembly consists of four contigs with a total genome size of  $\approx$ 7.85 Mbps. Approximately 93% of the genome is contained within two large contigs (4.2 and 3.1 Mbps). Remaining sequences were divided into two smaller contigs ranging from 420 to 153 kbps. A total of 7,247 candidate protein-coding genes were predicted using RAST (13) with a total G+C content of 63.5%. Upon comparison with genomes available within the RAST the closest relative to *Janthinobacterium* sp. CG23\_2 was determined to be *Janthinobacterium* sp. Marseille (score 542).

**Nucleotide sequence accession numbers.** This genome sequence has been deposited in EMBL/GenBank under the accession number [CYSS000000000](https://www.ncbi.nlm.nih.gov/nuccore/CYSS000000000). The version described in this paper is the first version, CYSS000000001.

## ACKNOWLEDGMENTS

Funding for this research came from the National Science Foundation (OPP-0838970 and 1141978), and the Department of Microbiology at Montana State University. H.J.S. was supported by the NASA Earth and Space Science Fellowship (NESSF) program.

Logistical support was provided by Raytheon Polar Services and Petroleum Helicopters Incorporated.

Any opinions, findings, and conclusions or recommendations expressed in this do not necessarily reflect the views of the National Science Foundation.

## FUNDING INFORMATION

NASA Earth and Space Science Fellowship provided funding to Heidi J. Smith. National Science Foundation (NSF) provided funding to Christine M. Foreman under grant number OPP-0838970. National Science Foundation (NSF) provided funding to Christine M. Foreman under grant number ANT-1141978.

## REFERENCES

1. Staehelin J, Harris NRP, Appenzeller C, Eberhard J. 2001. Ozone trends: a review. *Rev Geophys* 39:231–290. <http://dx.doi.org/10.1029/1999RG000059>.
2. Nienow JA, McKay CP, Friedmann EI. 1988. The cryptoendolithic microbial environment in the Ross Desert of Antarctica: light in the photo-synthetically active region. *Microb Ecol* 16:271–289. <http://dx.doi.org/10.1007/BF02011700>.
3. Dieser M, Greenwood M, Foreman CM. 2010. Carotenoid pigmentation in Antarctic heterotrophic bacteria as a strategy to withstand environmental stresses. *Arctic Antarct Alpine Res* 42:396–405. <http://dx.doi.org/10.1657/1938-4246-42.4.396>.
4. Mojib N, Farhoomand A, Andersen DT, Bej AK. 2013. UV and cold tolerance of a pigment-producing Antarctic *Janthinobacterium* sp. Ant5-2. *Extremophiles* 17:367–378. <http://dx.doi.org/10.1007/s00792-013-0525-9>.
5. Durán N, Menck CFM. 2001. *Chromobacterium violaceum*: a review of pharmacological and Industrial perspectives. *Crit Rev Microbiol* 27: 201–222. <http://dx.doi.org/10.1080/20014091096747>.
6. Konzen M, De Marco D, Cordova CAS, Vieira TO, Antônio RV, Creczynski-Pasa TB. 2006. Antioxidant properties of violacein: possible relation on its biological function. *Bioorg Med Chem* 14:8307–8313. <http://dx.doi.org/10.1016/j.bmc.2006.09.013>.
7. Kim SJ, Shin SC, Hong SG, Lee YM, Lee H, Lee J, Choi I-G, Park H. 2012. Genome sequence of *Janthinobacterium* sp. strain PAMC 25724, isolated from alpine glacier cryoconite. *J Bacteriol* 194:2096. <http://dx.doi.org/10.1128/JB.00096-12>.
8. Hornung C, Poehlein A, Haack FS, Schmidt M, Dierking K, Pohlen A, Schulenburg H, Blokesch M, Plener L, Jung K, Bonge A, Krohn-Molt I, Utpatel C, Timmermann G, Spieck E, Pommerening-Röser A, Bode E, Bode HB, Daniel R, Schmeisser C, Streit WR. 2013. The *Janthinobacterium* sp. HH01 genome encodes a homologue of the *V. cholerae* CqsA and *L. pneumophila* LqsA autoinducer synthases. *PLoS One* 8:e55045. <http://dx.doi.org/10.1371/journal.pone.0055045>.
9. Shoemaker WR, Muscarella ME, Lennon JT. 2015. Genome sequence of the soil bacterium *Janthinobacterium* sp. KBS0711. *Genome Announc* 3(3):e00689-15. <http://dx.doi.org/10.1128/genomeA.00689-15>.
10. Korlach J, Bjornson KP, Chaudhuri BP, Cicero RL, Flusberg BA, Gray JJ, Holden D, Saxena R, Wegener J, Turner SW. 2010. Real-time DNA sequencing from single polymerase molecules. *Methods Enzymol* 472: 431–455. [http://dx.doi.org/10.1016/S0076-6879\(10\)72001-2](http://dx.doi.org/10.1016/S0076-6879(10)72001-2).
11. Chin C, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Meth* 10:563–569. <http://dx.doi.org/10.1038/nmeth.2474>.
12. Koren S, Harhay GP, Smith TP, Bono JL, Harhay DM, McVey SD, Radune D, Bergman NH, Phillippy AM. 2013. Reducing assembly complexity of microbial genomes with single-molecule sequencing. *Genome Biol* 14:R101. <http://dx.doi.org/10.1186/gb-2013-14-9-r101>.
13. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.