

TEMPERATURE DEPENDENCY OF BIOLOGICAL DENITRIFICATION WITH ORGANIC MATERIALS ADDITION

ZBIGNIEW LEWANDOSWKI

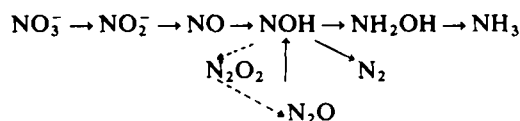
Polish Academy of Sciences, Institute of Environmental Engineering, 41-800 Zabrze, Skłodowskiej-Curie 34, Poland

(Received January 1981)

Abstract—The rate of a waste water denitrification process depends amongst others upon the temperature and the kind of compound used as electron donor. The results of measurements of denitrification kinetics using methanol, acetone and acetic acid have been presented.

INTRODUCTION

In recent years much attention has been given to biological denitrification. Many investigators have examined factors influencing denitrification sequence, i.e. anoxic conversion of nitrate through nitrite to elemental nitrogen gas. Dissimilatory nitrate metabolism occurs in a series of enzyme catalyzed reactions similar to the route hypothesized by Fewson & Nicholas (1961):



— enzymatic
 --- non-enzymatic.

The reaction sequence is partially confirmed through the enzymes found in denitrifying bacteria: nitrate reductase, nitrite reductase, nitric oxide reductase, hyponitrite reductase (not very well established), and hydroxylamine reductase (Christensen & Harremoës, 1972). In these reactions nitrate is reduced by serving as a hydrogen acceptor. An additional, but a relatively small quantity of nitrate is reduced to ammonia in order to supply nitrogen for cell synthesis. Typical denitrification genera listed by Painter (1970) are *Pseudomonas* sp., *Micrococcus* sp., *Spirillum* sp. and *Achromobacter* sp. In the absence of molecular oxygen these organisms use nitrate or nitrite as terminal electron acceptors while oxidizing organic matter for energy.

For application in wastewater treatment the denitrification process must operate over a wide range of temperatures. The variation in the denitrification rate, expressed in terms of the various temperature employed is given, according to Sutton *et al.* (1975), in Table 1. The denitrification rate depends not only on the temperature but amongst others the kind of compound utilized as electron donor. The effect of denitri-

fication by means of activated sludge at different temperatures using methanol, acetone and acetic acid as electron donors were examined.

KINETIC CONSTANTS

In non-limiting nutrient systems the denitrification reaction is the zero order. The reaction rate is given by:

$$k = \frac{dc}{dt} \quad (1)$$

where:

k = reaction rate, $\text{mg l}^{-1} \text{h}^{-1}$
 c = substrate concentration, mg l^{-1}
 t = time, h.

The effect of temperature on the reaction rate can be expressed by a temperature coefficient Q_{10} :

$$Q_{10} = \frac{k_{t+10}}{k_t} \quad (2)$$

where:

k_t = reaction rate at temperature t , $\text{mg l}^{-1} \text{h}^{-1}$
 k_{t+10} = reaction rate at temperature $t + 10$, $\text{mg l}^{-1} \text{h}^{-1}$.

The value of Q_{10} is only approximate because it depends on the range of temperatures over which the measurement was done. Reaction rate is usually evaluated at 20°C. The relationship between the measured values and the reaction rate at 20°C is:

$$k_{20} = \frac{k_t}{\theta^{t-20}} \quad (3)$$

where:

θ = constant.

The temperature effect on the reaction rate is indicated by the value of the activation energy in the Arrhenius equation:

$$k = A \cdot e^{-E_a/RT} \quad (4)$$

Table 1. Temperature coefficients and denitrification (Sutton *et al.*, 1975)

	Temperature range (°C)	Arrhenius activation energy ($E_a = \text{cal mol}^{-1}$)	θ Values	Q_{10} values ($Q_{10} = \theta^{10}$)	Reference
Activated sludge SRT = 6 days	6-25	15,900	1.09	2.5	Sutton (1973) Pilot plant
	6-16				
	10-20				
Upflow packed column	5-25	11,090	1.07	2.1	
	5-15				
	10-20				
Batch <i>P. denitrificans</i>	3-27	16,800	1.12	3.0	Dawson & Murphy (1972) Lab scale
	10-20				
Batch	15-25	10,000	1.06	1.74	Stensel (1970) Lab scale
Activated sludge SRT = 2 days Continuous					
activated sludge	10-20	19,500	1.13	3.3	Mulbarger <i>et al.</i> (1970) Pilot plant
Activated sludge SRT = 7.6 days	10-20	19,000	1.15	3.3	
Activated sludge	10-20			2	
Batch	10-20			2.6	Wuhrman & Mechsner (1965) Lab scale
Activated sludge					

where:

A = frequency factor

E_a = activation energy, cal mol^{-1}

R = universal gas constant, $\text{cal mol}^{-1} \text{deg}^{-1}$

T = absolute temperature, K.

With the use of a logarithmic transformation we could obtain a linearized form of the Arrhenius equation:

$$\ln k = \ln A - \frac{E_a}{RT} \quad (5)$$

This model, using decimal logarithmic scale, was fitted to the experimental data using the least squares fit. Calculation of the slope for the linearized model provided an evaluation of the Arrhenius activation energy:

$$E_a = \frac{4.576 \cdot T_1 \cdot T_2}{T_1 - T_2} \log \frac{k_1}{k_2} \quad (6)$$

As the reaction rate in biological reactors depends on the microorganisms concentration it was taken into consideration that the relative specific reaction rate is:

$$k' = \frac{k}{X} \quad (7)$$

where:

k' = relative specific reaction rate, h^{-1}

k = measured reaction rate, $\text{mg l}^{-1} \text{h}^{-1}$

X = concentration of microorganisms, mg l^{-1} .

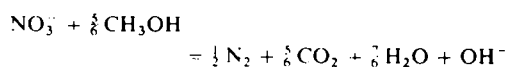
EXPERIMENTAL PROCEDURE

Fresh activated sludge was obtained from the Vienna-Blumental Sewage Treatment Plant which works according

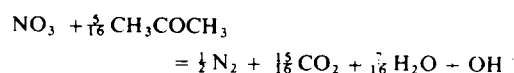
to the simultaneous nitrification-denitrification process (Matsche, 1977). In the treatment plant wastewater of about 200,000 PE is treated without primary sedimentation in aeration tanks with mammoth rotor aeration.

Generally in each experiment the activated sludge was transferred into 1 l. plastic containers which were placed in a thermostatic bath. Complete mixing was accomplished by means of magnetic stirrer. The experiments were carried out at 2.5, 5, 10, 15, 20, 25, 30 and 35°C. Before the measurement the samples were moderately aerated and adapted to the temperature of measurement for 2 h. After 2 h a solution of $10 \text{ g l}^{-1} \text{KNO}_3$ was added to each of the samples to obtain a concentration of about $30 \text{ mg l}^{-1} \text{N-NO}_3$ within the liquor. The nitrate concentration was measured by means of ion-selective electrode. Next, the solution of methanol, acetone and acetic acid were added respectively. The stoichiometric reactions during denitrification are as follows:

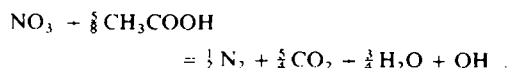
with addition of methanol:



with addition of acetone:



with addition of acetic acid:



The concentration of the added reagents were respectively: methanol 75 mg l^{-1} , acetone 65 mg l^{-1} , acetic acid 85 mg l^{-1} within the liquor. Aeration was stopped and the oxygen consumption was observed by means of oxygen electrode. The measurement began 5 min after obtaining the anoxic conditions and the samples were collected every 30 min for 3 h in the case of low temperatures and every 10 min for 1 h at higher temperatures. Samples were filtered through filter paper (Schleicher und Schüll No. 595) and the concentration of N-NO_3 was measured by use of a Technicon analyzer. The concentration of microorganisms was measured as MLSS.

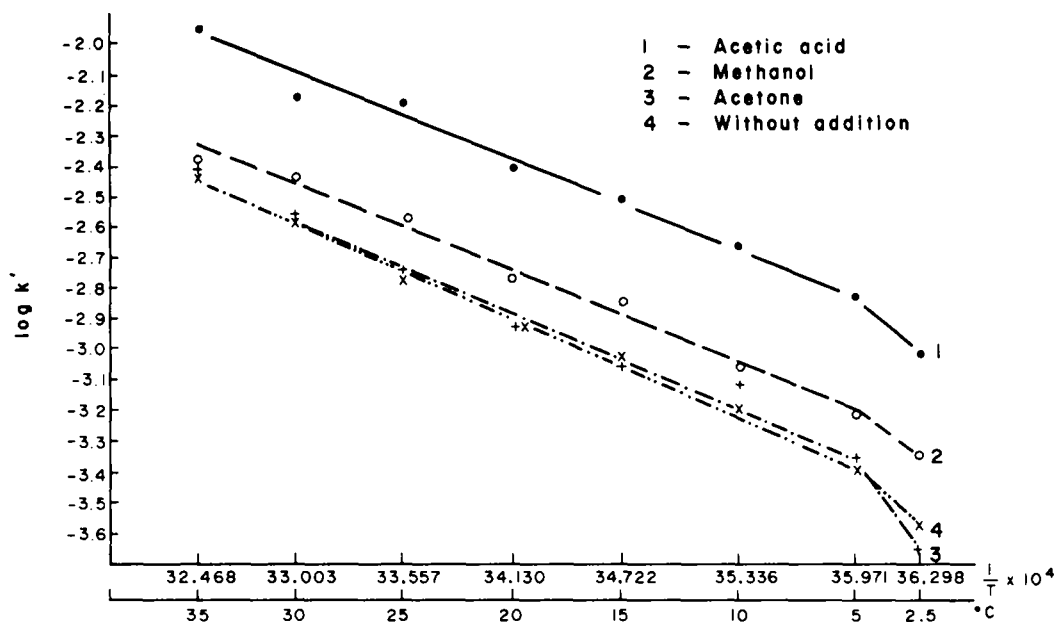


Fig. 1. Arrhenius dependence of rate constant for denitrification with addition of synthetic carbon source.

RESULTS

Figure 1 shows the results of a study. From these data the kinetic constants were found and presented in Table 2.

The data shows that the relationship between specific reaction rate and temperature within the range 5–35°C is linear. Below 5°C, the slowdown in reaction rate is greater than could be expected on the basis of results obtained in the range 5–35°C. The least squares fit for the data in this range is as follows:

With addition of methanol:

$$\log k' = -2422 \frac{1}{T} + 5.5288 \quad r = 0.9927$$

With addition of acetone:

$$\log k' = -2602 \frac{1}{T} + 6.0109 \quad r = 0.9871$$

With addition of acetic acid:

$$\log k' = -2369 \frac{1}{T} + 5.7083 \quad r = 0.9906$$

Without any addition:

$$\log k' = -2677 \frac{1}{T} + 6.2472 \quad r = 0.9948.$$

The sample without any organic chemical addition shows relatively high denitrification rate. This is probably caused by the type of plant used at the Vienna-Blumental Treatment Plant. The Plant works without primary sedimentation. The wastewater after removal of coarse material by bar screens and withdrawal of sand and floatable material in an aerated grit chamber, flows directly to the aeration tanks which no doubt results in more than the normal amount of organic matter being adsorbed onto the flocs and so causing the high rate of exogenous denitrification.

Acetic acid was most effective at increasing the denitrification rate followed by methanol, whilst the use of acetone showed no improvement over the undosed sample. The scale of improvement can be shown by simple comparison. The denitrification rate with addition of acetic acid at 5°C is approximately the same as that at 23°C without any addition.

Table 2. Kinetic constants

Chemical	$k'_{20} \cdot 10^3$ (h ⁻¹)	$k'_{10} \cdot 10^3$ (h ⁻¹)	Q_{10}	θ	
				$\theta = 10^{\sqrt{Q_{10}-1}}$	E_a (cal mol ⁻¹)
Without addition	1.29	0.61	2.11	1.08	12,300
Methanol	1.83	0.93	1.97	1.07	11,200
Acetone	1.35	0.66	2.05	1.07	11,800
Acetic acid	4.20	2.17	1.94	1.07	10,900

It is well known that the relationship between the specific reaction rate and the temperature is, in case of the enzyme catalyzed reactions, linear only in a relatively narrow range. The results which were obtained show that the lower point of inflexion is below 5°C and does not depend on the kind of organic compound used for improving the amount of denitification achieved.

The Arrhenius activation energy E_a was in all cases more or less the same and varied in the range 10.9–12.3 kcal mol⁻¹. Parameter Q_{10} changes in the range 1.94–2.11 and does not depend on the type of organic materials consumed. This shows evidently that in all cases the reaction course according to the van't Hoff rule. The comparison of obtained results with the data presented in Table 1 shows that the kinetic constants like Q_{10} , θ and E_a only vary slightly, and lie about the theoretical values obtained from the van't Hoff rule.

Acknowledgements—This study is abstracted in part from work supported by research grant from Austrian Bundesministerium für Wissenschaft und Forschung. The grant was sponsored by UNESCO. The author is grateful to Professor Dr ing. Wilhelm von der Emde and Dr Norbert Matsche from Institute für Wasserversorgung, Abwasserreinigung und Gewässerschutz der Technischen Universität Wien for their help and advice.

REFERENCES

Christensen M. H. & Harremoës P. (1972) Biological denitrification in water treatment. A literature study. Depart-

ment of Sanitary Engineering. Technical University of Denmark. Report 2-72.

Dawson R. N. & Murphy K. L. (1972) The temperature dependency of biological denitrification. *Water Res.* **6**, 71.

Fewson C. A. & Nicholas D. J. D. (1961) Utilization of nitrate by microorganisms. *Nature* **190**, 2-7.

Johnson W. K. & Vania G. B. (1971) Nitrification and denitrification of waste water. Sanitary Engineering Report No 1755, Universities of Minneapolis. Minneapolis—St Paul.

Matsche N. F. (1977) Removal of nitrogen by simultaneous nitrification-denitrification in an activated sludge plant with Mammoth rotor aeration. *Prog. Wat. Technol.* **8**, 625-637.

Matsche N. F. & Spatzierer G. (1977) Investigation towards a control of simultaneous nitrogen elimination in the treatment plant Vienna-Blumental. *Prog. Wat. Technol.* **8**, 501-508.

Mulbarger M. C. *et al.* (1970) Modifications of the activated sludge process for nitrification and denitrification. Paper presented at 43 Annual Conference, Water Pollution Control Federation, Boston, MA.

Painter H. A. (1970) A review of literature on inorganic nitrogen metabolism in microorganisms. *Water Res.* **4**, 393-450.

Stensel H. D. (1970) Biological kinetics of the suspended growth denitrification process. Ph. D. Thesis, Cornell University Ithaca, NY.

Sutton P. M. (1973) Continuous biological denitrification of wastewater. M.Sc. Thesis, Department of Chemical Engineering, McMaster University of Hamilton, Ontario, Canada.

Sutton P. M. *et al.* (1975) Low-temperature biological denitrification of wastewater. *J. Wat. Pollut. Control. Fed.* **47**, 122-134.

Wuhrman K. & Mechsner K. (1965) Beitrag zur Kenntnis der Mikrobiellen Denitrifikation. *Path. Microbiol.* **26**, 579.